

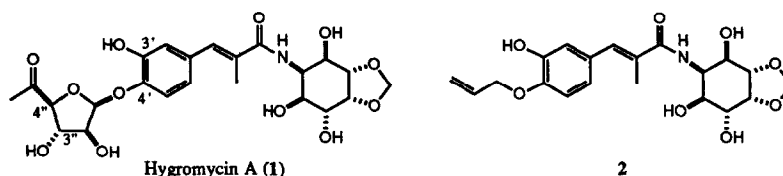
SYNTHESIS AND *IN VITRO* ANTIBACTERIAL ACTIVITY OF HYGROMYCIN A ANALOGS MODIFIED AT THE C_{4'} ARYL POSITION

Burton H. Jaynes,* Christopher B. Cooper, Scott J. Hecker, Kyle T. Blair, Nancy C. Elliott,
Susan C. Lilley, Martha L. Minich, Douglas L. Schicho, and Kim M. Werner
Pfizer Inc, Central Research Division, Groton, CT 06340

(Received in USA 25 March 1993)

Abstract: A variety of hygromycin A analogs are described which contain modifications of or replacements for the natural sugar. Antibacterial activities against *Serpulina (Treponema) hyodysenteriae*, an important animal health pathogen, are reported and indicate that small lipophilic C_{4'} substituents serve as useful sugar surrogates in the hygromycin A class.

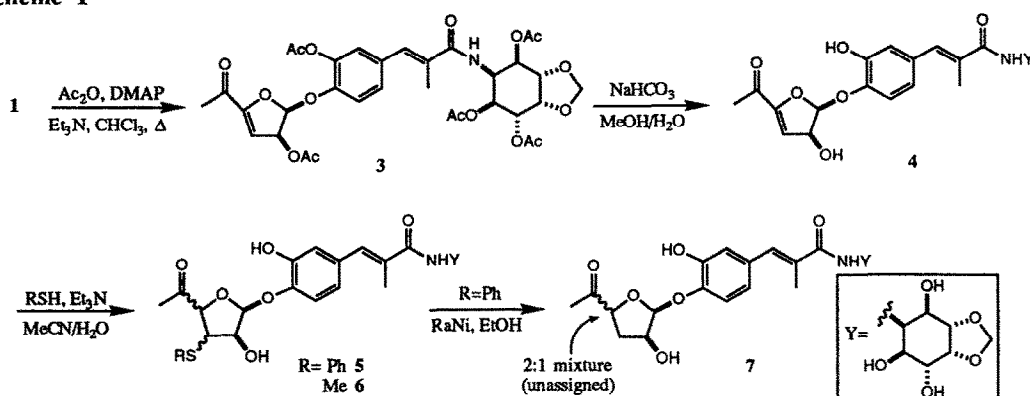
Hygromycin A (1) is a fermentation-derived natural product first isolated from *Streptomyces hygroscopicus* in 1953.¹ Although known to possess moderate antibacterial activity, it was only recently discovered to have excellent *in vitro* potency² against *Serpulina (Treponema) hyodysenteriae*, the causative agent of swine dysentery, an economically significant muco-hemorrhagic disease of swine. More importantly, the antibiotic also demonstrated efficacy in the treatment of induced dysentery infection in pigs at levels of 5-20 g/ton feed.³ The renewed interest in hygromycin A has led to increased synthesis activities, including a recent total synthesis.⁴ Additional reports from our laboratories have detailed structure-activity relationships (SAR) about the aromatic ring,⁵ the enamide,⁶ and the aminocyclitol.⁷ In addition, we described the remarkable functional equivalence of simple lipophilic alkyl ethers in place of the *arabino*-furanosyl aryl substituent of hygromycin A, represented by allyl ether 2.⁸ This paper further describes the range of accepted groups at the C_{4'} position, including modifications of the natural sugar, and oxygen, nitrogen, and carbon-linked substituents.



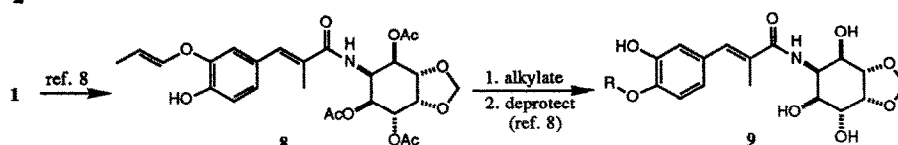
Initial semisynthetic studies on hygromycin A's furanose moiety suggested that this region's SAR may be very sensitive to minor alterations in structure. By acetylation of the natural product with concomitant elimination, enone 3 is produced (Scheme 1); acetate cleavage with wet NaHCO₃/MeOH affords dihydrofuran 4 which retains only moderate *in vitro* antibacterial activity⁹ (*S. hyodysenteriae* MIC = 25 µg/mL). In order to reintroduce sp³ hybridization at C_{3'} and C_{4'}, phenyl and methyl thiolate anions were added in a conjugate fashion but failed to restore *in vitro* activity¹⁰ (MICs of 5 and 6: >100 µg/mL). Subsequent desulfurization was accomplished with Raney nickel to provide an inactive mixture of the 3''-deshydroxy hygromycin A epimers 7¹⁰ (MIC of 7: >100 µg/mL), indicating that inactivity of the sulfides is not due to simple steric repulsion at C_{3'}, and implicating an absolute requirement for the C_{3'} hydroxyl group.

With this surprising stringency uncovered in the furanose region, we set about to determine if complete replacement of the *arabino*-furanose, patterned after successful efforts in the antiviral acyclic nucleoside field,¹² would yield analogs less sensitive to variation. We sought an acyclic sugar surrogate which would alleviate potential C_{4'} epimerization problems (hygromycin A is isolated as a mixture of epimers) and provide the minimum structural requirements for antibacterial activity. Methodology was developed to cleave the sugar of hygromycin A and to derivatize selectively the resultant C_{4'} phenolic hydroxyl group.⁸ Through intermediate **8**, a variety of ethers were prepared by halide displacement or Mitsunobu reaction with an alcohol, followed by deprotection (Scheme 2). *In vitro* activities of some of these analogs are listed in Table 1.

Scheme 1

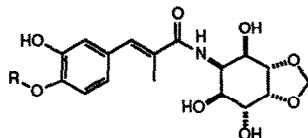


Scheme 2



The *in vitro* results in Table 1 suggest that small lipophilic groups (less than five methylene groups, in general) lead to the most active C_{4'} substituted analogs. (And this has been shown to translate to *in vivo* activity in both mice⁸ and swine.¹³) Analogs bearing longer aliphatic chains at C_{4'} are generally less potent (**9k**, **9l**), although activity may be restored through the introduction of polar groups to the sidechain (compare pentyl ether **9l** with methyl sulfide **9o** and ketone **9p**). Hydroxyl groups, designed to overlay with those at C_{2'} and C_{3'} of the natural furanose, do not improve potencies (entries **9t**, **9u**, **9v**). Steric hindrance influences antibacterial activity as branched chain substituents decrease potency (**2** versus **9g** and **9h**) while cyclic structures bearing a greater number of carbon atoms retain activity (**9m**, **9q**, **9r**). Interestingly, "typical" antiviral sidechains¹² (**9s**, **9t**, **9u**, **9v**) do not serve as effective sugar surrogates for hygromycin A and activity actually *decreases* as hydroxyl groups are added, a remarkable finding considering the parent structure's hydrophilic furanose.

Table 1. 4' ether analogs



Compound	R	<i>S. hyodysenteriae</i> MIC (μg/mL)	Method of preparation
Hygromycin A (1)	---	0.78-1.56	---
2	CH ₂ =CHCH ₂	0.78	ref. 14, 15
9a	OR = H	>200	ref. 16
9b	H	200	ref. 15, 17
9c	CH ₃	12.5	ref. 18
9d	CH ₃ CH ₂	1.56	ref. 18
9e	FCH ₂ CH ₂	6.25	ref. 19
9f	CH ₃ CH ₂ CH ₂	0.78-1.56	ref. 18
9g	CH ₂ =CHCH(CH ₃)	25	ref. 19
9h	CH ₂ =C(CH ₃)CH ₂	3.13	ref. 19
9i	propargyl	6.25	ref. 19
9j	cyclopropylCH ₂	0.39-0.78	ref. 19
9k	CH ₃ CH ₂ CH ₂ CH ₂	1.56	ref. 18
9l	CH ₃ CH ₂ CH ₂ CH ₂ CH ₂	>200	ref. 18
9m	cyclopentyl	6.25	ref. 19
9n	NH ₂ COCH ₂	>200	ref. 20
9o	CH ₃ SCH ₂ CH ₂ CH ₂	0.78-1.56	ref. 19
9p	CH ₃ (CO)CH ₂ CH ₂ CH ₂	12.5-25	ref. 19
9q	PhCH ₂	25	ref. 19
9r	thiopheneCH ₂	3.13-6.25	ref. 19
9s	CH ₃ CH ₂ OCH ₂	25	ref. 21
9t	HOCH ₂ CH ₂	50-100	ref. 22
9u	HOCH ₂ CH ₂ OCH ₂	25	ref. 14, 25
9v	HOCH ₂ CH(OH)CH ₂	25	ref. 26

To study the importance of the linking ether oxygen at C_{4'}, several nitrogen and carbon-linked analogs were prepared. In general, these analogs were synthesized from the appropriately substituted benzaldehyde by olefination using a three-step sequence initiated with a stabilized Wittig reaction or a one-step phosphonate procedure (Scheme 3).⁵ The surprising tolerance for simple lipophilic side chains is also observed when the oxygen at C_{4'} is replaced with nitrogen or carbon as seen in examples from Table 2. Although the MICs are not as good as in the oxygen-linked series, similar trends are observed with the various lipophilic substituents.

Previously published work described the tolerance of the C_{3'} position for proton, amino and fluoro substitution in addition to the natural hydroxy functionality.⁵ Indeed, the C_{4'} structure-activity relationships are maintained when the 3-position hydroxyl group is replaced with a hydrogen (13g, 13k, and 13l) or fluorine atom (13h and Table 3) in the C_{4'} oxygen, nitrogen, and carbon-linked series.

Scheme 3

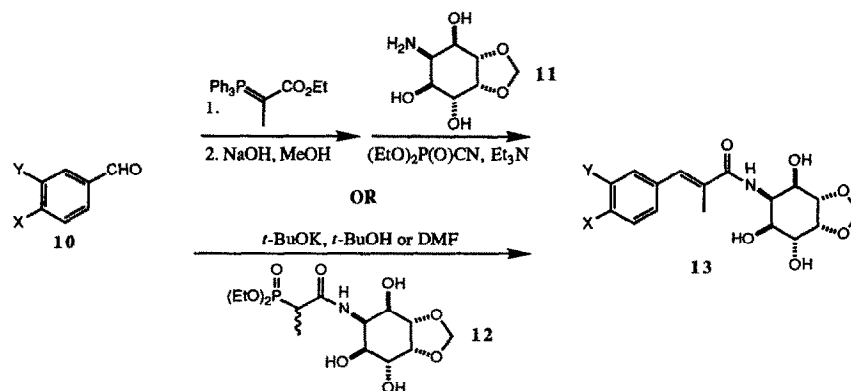
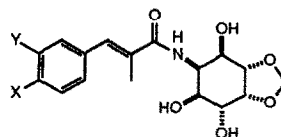


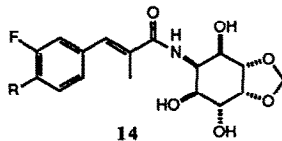
Table 2. 4' amino and carbon analogs



Compound	X	Y	<i>S. hyodysenteriae</i> MIC ($\mu\text{g/mL}$)	Method of preparation
13a	NH_2	OH	>200	ref. 27
13b	$\text{CH}_3\text{CH}_2\text{NH}$	OH	100	ref. 29
13c	$\text{CH}_3\text{CH}_2\text{CH}_2\text{NH}$	OH	12.5-25	ref. 29
13d	$\text{CH}_2=\text{CHCH}_2\text{NH}$	OH	12.5-25	ref. 29
13e	$\text{CH}_3\text{CH}_2\text{CH}_2(\text{Et})\text{N}$	OH	>200	ref. 29
13f	$\text{CH}_2=\text{CHCH}_2(\text{Ac})\text{N}$	OH	>200	ref. 30
13g	pyrrolidin-1-yl	H	6.25	ref. 16, 31
13h	pyrrolidin-1-yl	F	6.25	ref. 16, 31
13i	$\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}_2$	OH	3.13-6.25	ref. 16, 32
13j	PhCH_2	OH	>200	ref. 16
13k	$\text{CH}_3\text{CH}_2\text{CH}_2\text{CH}_2$	H	12.5-25	ref. 16, 32
13l	Ph	H	12.5	ref. 16

In summary, despite dramatic differences from the natural furanose of hygromycin A, analogs possessing small lipophilic C_4' substituents, whether linked through an oxygen (9b-v, 14a-c), nitrogen (13a-h), or carbon atom (13i-l), maintain comparable *in vitro* activity relative to the natural product as measured against *S. hyodysenteriae*. Several of these *in vitro* actives have been shown to possess *in vivo* activity similar to hygromycin A in mouse and swine *S. hyodysenteriae* infection models.^{8,13} In contrast, analogs incorporating hydroxyl groups on the C_4' side chain designed to overlay with the natural furanose alcohols, and thus bind similarly to a molecular target,³⁴ fail to retain *in vitro* antibacterial activity. In fact, minor modifications that employ the furanose as a template for semisynthesis lead to a decrease in potency.

Table 3. 3' fluoro analogs



Compound	R	<i>S. hyodysenteriae</i> MIC (μg/mL)	Method of preparation
14a	CH ₃ O	6.25-12.5	ref. 16, 33
14b	CH ₂ =CHCH ₂ O	1.56	ref. 5
14c	propargyl-O	6.25	ref. 16, 33
14d	F	>200	ref. 16

Acknowledgements: We wish to thank Mr. W.P. Cullen and Mr. J.R. Oscarson for generous supplies of hygromycin A from fermentation, and Mr. S.B. Seibel for microbiological testing.

References and Notes:

- Pittenger, R.C.; Wolfe, R.N.; Hoehn, M.M.; Marks, P.N.; Daily, W.A.; McGuire, J.M. *Antibiot. Chemother.* **1953**, *3*, 1268; Mann, R.L.; Gale, R.M.; van Abeele, F.R. *Ibid.* **1953**, *3*, 1279.
- Omura, S.; Nakagawa, A.; Fujimoto, T.; Saito, K.; Otoguro, K.; Walsh, J.C. *J. Antibiot.* **1987**, *40*, 1619.
- Nakagawa, A.; Fujimoto, T.; Omura, S.; Walsh, J.C.; Stotish, R.L.; George, B. *J. Antibiot.* **1987**, *40*, 1627.
- Chida, N.; Ohtsuka, M.; Nakazawa, K.; Ogawa, S. *J. Org. Chem.* **1991**, *56*, 2976.
- Hecker, S.J.; Cooper, C.B.; Blair, K.T.; Lilley, S.C.; Minich, M.L.; Werner, K.M. *BioMed. Chem. Lett.* **1993**, *3*, 289.
- Hecker, S.J.; Minich, M.L.; Werner, K.M. *BioMed. Chem. Lett.* **1992**, *2*, 533.
- Hecker, S.J.; Lilley, S.C.; Minich, M.L.; Werner, K.M. *BioMed. Chem. Lett.* **1992**, *2*, 1015; Hecker, S.J.; Lilley, S.C.; Werner, K.M. *Ibid.* **1992**, *2*, 1043.
- Jaynes, B.H.; Elliott, N.C.; Schicho, D.L. *J. Antibiot.* **1992**, *45*, 1705.
- Hygromycin A MIC (minimum inhibitory concentration) = 0.78-1.56 μg/mL. *S. hyodysenteriae* MICs were determined as in Weber, F.H.; Earley, D.L. *Antimicrob. Agents Chemother.* **1991**, *35*, 2012.
- All possible diastereomers were obtained for analogs 5, 6, and 7, but not separated; the known C₄" epimer of hygromycin,¹¹ epihygromycin A, has an MIC against *S. hyodysenteriae* of 6.25 μg/mL.
- Wakisaka, Y.; Koizumi, K.; Nishimoto, Y.; Kobayashi, M.; Tsuji, N. *J. Antibiot.* **1980**, *33*, 695.
- For example, see: Chu, C.K.; Cutler, S.J. *J. Heterocyclic Chem.* **1986**, *23*, 289.
- Illyes, E.F.; Kamicker, B.J.; Jaynes, B.H.; LeMay, M.A.; Shively, J.E., manuscript in preparation.
- Prepared as outlined in reference 8, by alkylation with the appropriate bromide.
- An improved procedure for the preparation of hygromycin A aglycone (**9b**) and allyl ether analog **2** will be reported elsewhere.
- Prepared from the appropriately substituted benzaldehyde as illustrated in Scheme 3.⁵

17. Mann, R.L.; Woolf, D.O. *J. Am. Chem. Soc.* **1957**, *79*, 120.
18. Prepared as outlined in reference 8, by alkylation with the appropriate iodide.
19. Prepared as outlined in reference 8, by Mitsunobu reaction with the appropriate alcohol.
20. Prepared as outlined in reference 8, by Mitsunobu reaction with methyl glycolate. In the acetate removal step (NH₃/MeOH), the methyl ester was converted to primary amide 9n.
21. Prepared as outlined in reference 8, by alkylation of the protected aglycone with chloromethyl ethyl ether and N,N-diisopropylethylamine.
22. Compound 9f was prepared from 3-(allyloxy)-4-hydroxybenzaldehyde²³ by (1) ethylene sulfate opening with the sodium anion (NaH, THF) followed by *in situ* sulfate hydrolysis,²⁴ (2) olefination with (carboxyethylidene)triphenylphosphorane in CH₂Cl₂, (3) allyl group removal,⁸ (4) ester saponification (aq. NaOH, MeOH), (5) hydroxyl group protection with Ac₂O, DMAP, and Et₃N, (6) coupling with aminocyclitol 11 ((EtO)₂P(O)CN and Et₃N in DMF), and (7) deacetylation (K₂CO₃, MeOH).
23. Reitz, A.; Avery, M.A.; Verlander, M.S.; Goodman, M. *J. Org. Chem.* **1981**, *46*, 4859.
24. Gao, Y.; Sharpless, K.B. *J. Am. Chem. Soc.* **1988**, *110*, 7538; Kim, B.M.; Sharpless, K.B. *Tetrahedron Lett.* **1989**, *30*, 655.
25. (2-Acetoxyethoxy)methyl bromide was prepared from acetyl bromide and 1,3-dioxolane: Robins, M.J.; Hatfield, P.W. *Can. J. Chem.* **1982**, *60*, 547.
26. Compound 9v was prepared from 4-(allyloxy)-3-hydroxybenzaldehyde²³ by (1) catalytic *cis*-hydroxylation (OsO₄, NMO, THF, H₂O), (2) olefination with (carboxyethylidene)-triphenylphosphorane in THF, (3) ester saponification (aq. NaOH, MeOH), (4) hydroxyl group protection with Ac₂O, DMAP, and Et₃N, (5) coupling with aminocyclitol 11 ((EtO)₂P(O)CN and Et₃N in DMF), and (6) deacetylation (K₂CO₃, MeOH).
27. Compound 13a was prepared from 3-hydroxy-4-nitrobenzaldehyde by (1) allylation (allyl bromide, K₂CO₃, DMF), (2) olefination with (carboxyethylidene)triphenylphosphorane in CH₂Cl₂, (3) reduction to the aniline (SnCl₂, HCl), (4) allyl isomerization (Ir(COD)(MePh₂P)₂PF₆, THF),²⁸ (4) ester saponification (KOH, EtOH), (5) coupling with aminocyclitol 11 (EEDQ, DMF), and (6) enol ether hydrolysis (TFA, CH₂Cl₂, MeOH).
28. Oltvoort, J.J.; van Boeckel, C.A.A.; de Koning, J.H.; van Boom, J.H. *Synthesis* **1981**, 305.
29. Analogs 13b-e were prepared in a manner identical to that used for 13a with an alkylation step immediately before or after allyl isomerization.
30. Compound 13f was prepared from 13d by (1) peracetylation (Ac₂O, DMAP, Et₃N) and (2) de-*O*-acetylation (NaHCO₃, aq. MeOH).
31. 4-Pyrrolidinobenzaldehyde and 3-fluoro-4-pyrrolidinobenzaldehyde were prepared by reaction of pyrrolidine with 4-fluorobenzaldehyde and 3,4-difluorobenzaldehyde, respectively.
32. 4-Butyl-3-hydroxybenzaldehyde was prepared from vanillin by (1) triflate formation ((Tf)₂O, pyridine), Stille coupling (Bu₄Sn, (CH₃CN)₂PdCl₂, DMF), and (3) demethylation (BBr₃, CH₂Cl₂). 4-Butylbenzaldehyde was prepared similarly from 4-hydroxybenzaldehyde.
33. 3-Fluoro-4-methoxybenzaldehyde and 3-fluoro-4-propargyloxybenzaldehyde were prepared by reaction of sodium methoxide and sodium propargyloxide, respectively, with 3,4-difluorobenzaldehyde.
34. Mode of action studies have revealed a ribosomal peptidyltransferase inhibition mechanism for hygromycin A: Guerrero, M. del C.; Modolell, J. *Eur. J. Biochem.* **1980**, *107*, 409.